



Efficacy of nanocapsules loaded with *Ectomyelois ceratoniae* (Zeller) (Lepidoptera: Pyralidae) sex pheromone as evaluated in wind tunnel and field trapping experiments

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Abstract

The carob moth, *Ectomyelois ceratoniae* (Zeller) (Lep.: Pyralidae), is a worldwide pest of pomegranate. Although synthetic sex pheromone has been used extensively for the management of this pest, the major component (*Z,E*)-9,11,13-tetradecatrienal (trienal) is unstable. We microencapsulated synthetic trienal and demonstrated that a nanocapsule gel formulation was superior to unformulated trienal and other formulations, including the pheromone analogue (*Z,E*)-7,9,11-dodecatrienyl formate, in wind tunnel and field trapping experiments. No significant effect was observed between the total times spent by males from the release site to dispensers in different treatments in wind tunnels. In the field, the trienal nanocapsule formulation provided superior attraction for up to 4 weeks, while no other formulation induced trap catches for longer than 3 weeks. According to our finding, microencapsulation of *E. ceratoniae* pheromone components may provide improved trap lures and may be suitable for application in mating disruption.

Keywords Carob moth · pheromone · Microencapsulation · Pomegranate · (*Z, E*)-9,11,13-tetradecatrienal

Introduction

Pomegranate, *Punica granatum* L., (Myrtales: Punicaceae) originated from central Asia (Holland et al. 2009) and is now a cultivated crop species in many tropical and subtropical regions of the world, particularly India, Iran and Turkey (Kahramanoglu and Usanmaz 2016). In Iran, > 80,000 hectares of pomegranate orchards produce about one million tons of pomegranates each year (Ahmadi et al. 2016).

Carob moth, *Ectomyelois ceratoniae* (Zeller) (Lepidoptera: Pyralidae), is the main pest of pomegranate (Karami et al. 2010), date (Nay et al. 2006), almond (Madge 2014) crops worldwide, and in the Middle East causes 30 – 80% annual crop loss (Shakeri 2004; Mamay et al. 2016). Carob moth females lay their eggs on stamens in the calyx and the

larvae feed on petals, pierce the fruit and enter it, creating brown spots on the peel, as well as hollow, cracked and rotten fruit that appears black and moldy and is unmarketable (Mamay et al. 2014). Once the larvae penetrate the fruit chemical pesticides are ineffective (Shakeri 2004). In Iran's Lorestan Province, carob moth is tri-voltine, with infestations persisting from the end of April to mid-November (Naserian et al. 2013).

Control tactics used against the carob moth in Iran include collection and removal of overwintering larvae in rotten pomegranates (Sheikh Ali et al. 2009), stamen removal, application of kaolin clay powder to disrupt oviposition (Mazhab et al. 2014), covering the crown with netting (Rafeie et al. 2011), release of the egg parasitoid *Trichogramma embryophagum* (Hartig) (Rezaei-Azqandi et al. 2015) and use of pheromone mediated mating disruption (Tamhankar et al. 2000; Cichón et al. 2004) or mass trapping (Pezhman and Saeidi 2018).

The main components of *E. ceratoniae* sex pheromone are (*Z,E*)-9,11,13-tetradecatrienal (trienal), (*Z,E*)-9,11-tetradecadienal and (*Z*)-9-tetradecenal (Noorbakhsh et al. 2017a; Varshovi et al. 2018). Instability of trienal leads to low efficiency of mating disruption (Baker et al. 1991). A pheromone analogue, (*Z, E*)-7,9,11-dodecatrienyl formate,

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is only weakly attractive (Avand-Faghieh et al. 2012; Dhoubi et al. 2016). Successful pheromone applications require sustained release rates over the period that adult insects are active (Chen et al. 2018). Traditional formulations, such as microencapsulation, rubber septa, plastic tubes, hollow fibers and paraffin oil, can extend the duration of bioactivity (Gordon et al. 2005; Li et al., 2012). However, they provide inadequate control in field trials, and an alternative formulation is sought to retard breakdown of the trienal (Mitchell et al. 1976; Avand-Faghieh et al. 2012).

Recent advances in pest control technology include microencapsulation of chemical pesticides and pheromones (Racuciu et al. 2009; Roy et al. 2010; Heidary et al., 2020), employing materials such as polyethylene (Bradley et al. 1995), porous silica glass (Tiboni et al. 2008), zeolites (Muñoz-Pallares et al. 2001) and paraffin wax (Atterholt et al. 1999) to form nanocapsules, nanospheres, micelles, nanogels, liposomes and inorganic nano-cages (Nuruzzman et al. 2016; Chen et al. 2018) ranging in size from 10 to 1000 nm (Nagavarma et al. 2012). To achieve sustained release rates and efficacy, pheromones can be formulated into nanogels using a low molecular mass gelator (Bhagat et al. 2013), and hydrophobic pheromones can be encapsulated into nanoparticles (Chen et al. 2018; de Matos et al. 2019). Although many studies have been performed on the role of pheromones to control the carob moth, little is known about determining the efficacy of nanocapsule formulation of sex pheromone. Our objective was to evaluate the effect of nanoencapsulation on the stability and attractiveness of the trienal sex pheromone molecule as determined in wind tunnel and field experiments.

Materials and methods

Preparing the nanocapsule formulation

Analytical-grade materials used without further purifications were Gelatin (type G-2500) (MilliporeSigma, Merck KGaA, Darmstadt, Germany), gum Arabic (CARLO ERBA Reagents GmbH, Emmendingen, Germany), and thin layer chromatography (TLC) plates (25 TLC glass sheets 20×20, silica gel 60 F₂₅₄), hydrochloric acid and Tween 80 (Merck KGaA, Darmstadt, Germany). All aqueous solutions were prepared with distilled water. Synthesis of (*Z,E*)-9,11,13-tetradecatrienal (trienal) was performed according to Noorbakhsh et al. (2017b).

Nanoencapsulation was accomplished using complex coacervation (Sliwka 1975; Xing et al. 2005; Lv et al. 2013) of equal parts (w/v) of gelatin and gum Arabic (Danayetous et al. 2020). Firstly, 0.112 g of gelatin was dissolved in 7.5 mL of water making a 1.5 wt % solution. The solution was stirred for 0.5 h, heated to 50 °C, and 75 mg of

Tween 80 and 10 mg of the synthesized trienal were added while stirring. Then, 7.5 mL of the gum Arabic 1.5% solution was added, and the mixture was stirred at 200 rpm and sonicated for 1.0 h using a 250 W ultrasonicator (Ultrawave Ltd., Cardiff, UK). The pH of the solution was adjusted to 4.35 using diluted HCl and it was maintained at 50 °C for 1.0 h. Then, the nanocapsules were cross-linked by using formaldehyde (1 mL) at 40 °C. The resulting formulation was rinsed several times with distilled water, centrifugalized at 10,000 rpm for (15 min) and held in a vacuum oven at 30 °C for 12 h to obtain a white solid gel of nanoencapsulated trienal. Although the encapsulation efficiency was not measured chemically, the microcapsules mass product was washed with 1 mL dichloromethane and subjected to thin layer chromatography (hexane/ethyl acetate 4:1) in comparison with trienal, as seen in Fig. 1, three spots of trienal, mix trienal/ eluate microcapsules and elute microcapsules were placed on TLC and put it in TLC-tank with hexane/ethyl acetate eluent. After a few minute, the result showed the most trienal was adsorbed in microcapsules and no trienal was observed in eluate.

Morphology of nanocapsule formulation

The morphology of nanocapsule gel was characterized by depositing the suspension (2 µL) onto Formvar-coated

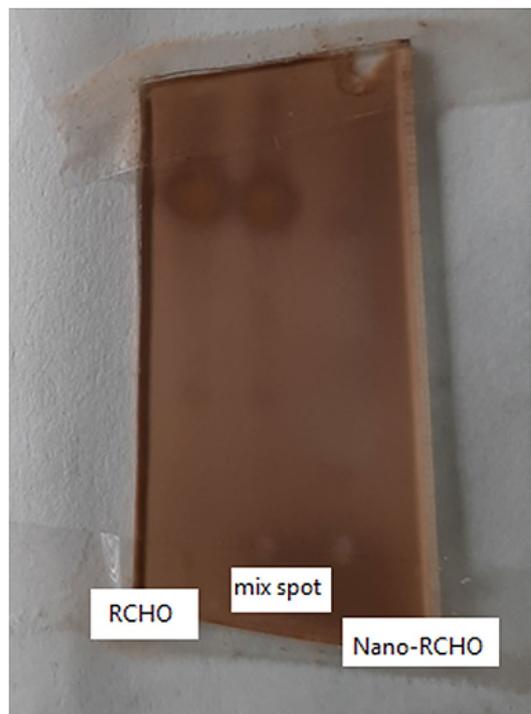


Fig. 1 TLC plate chromatography showing left spot, trienal particles; middle spot, mixture of unformulated trienal and trienal nanocapsules; right spot, trienal nanocapsules

copper grids (CM-300 Philips, Netherlands TEM), excess water was removed by filter paper, and the grids were allowed to dry for 1 h before examination by transmission electron microscopy (TEM) (200 kV Schottky field emitter HR) (Micro to Nano, Ltd., Haarlem, the Netherlands). The size of the nanocapsule particles was measured using purchased software (version 3.2, Soft Imaging System GmbH, Germany). The particle size and distribution of the nanocapsules were analyzed by a laser particle size analyzer (SZ-100z Dynamic Light Scattering and Zeta potential analyzer) (Horiba Ltd., Kyoto, Japan).

Ectomyelois ceratoniae rearing

Rotten pomegranates containing *E. ceratoniae* larvae were collected during November and December 2019 from the pomegranate orchards in western Lorestan Province, Iran. Larvae were excised and reared to maturity on fresh pomegranate fruits held in cages in incubators 29 ± 1 °C and $60 \pm 5\%$ RH. Emergent adults were held for 24 h and then used in experiments.

Wind tunnel experiment

Laboratory experiments were conducted in a wind tunnel ($50 \times 50 \times 150$ cm), wind speed 30 cm/s, red light (2 lx) held at 29 ± 1 °C and $60 \pm 10\%$ RH. The wind tunnel was kept in a room at 29 ± 1 °C, $60 \pm 10\%$ RH and under a photoperiod of 16L: 8 D h. To avoid odor contamination the room air was constantly renewed by an external fan. Four treatments, presented in randomized order, were 2.5 mg of synthetic trienal without dispenser in plastic vials, 1 mg of (*Z,E*)-7, 9, 11-dodecatrienyl formate on a rubber septum (Russell IPM, Deeside, UK), 100 mg of polymeric nanocapsule gel containing 2.5 mg of trienal) and control (no dispenser or pheromone). Adult males, 2–3 days old, were tested only once. In each bioassay, a single male was released 120 cm downwind of the test stimulus and given 3 min to initiate flight. Twenty males were tested against each stimulus. If no behavior was observed after 3 min, the moth was discarded. Sequential observations included time from release to take off, duration of upwind flight to 50 cm from the test stimulus, duration taken for close approach to within 10 cm of the test stimulus (Tasin et al. 2006; Borrero-Echeverry et al. 2015; Hatano et al. 2015; Celina et al. 2019), time resting (sitting) on the test stimulus and time taken in attempting to mate with the test stimulus.

Field experiment

A field experiment was conducted in pomegranate orchards, in Bizhenvand Village ($33^{\circ}41'16''$ N, $47^{\circ}50'43''$ E), Sarab-e-Dowreh County, Lorestan Province, Iran for 5 weeks from

September 7 to October 11, 2019. For all tests, Dispensers were placed in the middle of the sticky bottom (Daegil Co., Gyeongsangnam-do, South Korea) of white delta traps (200×200 base \times 100 mm high) (Raha Andish Kavan Co., Tehran, Iran). Traps were hung 150 cm over the ground, spaced 50 m apart within the canopy of pomegranate trees. Five treatments ($N = 4$ replicates) were applied in a randomized complete block design: 2.5 mg trienal in an Eppendorf tube, 1 mg (*Z, E*)-7, 9, 11-dodecatrienyl formate (Russell IPM, Deeside, UK), 100 mg of polymeric nanocapsule gel containing 2.5 mg of trienal, 100 mg if glycerin containing 2.5 mg of the trienal, and unbaited and control traps. Glycerin was tested as a potential alternative method to nanoencapsulation to improve stability of the trienal. Traps were checked weekly and captured males were counted and recorded.

Data analysis

Because there was no response to the control stimulus in the wind tunnel, controls were eliminated from analysis. The proportional responses to the three remaining treatments were analyzed with a chi-square test. Mean durations of behavioral categories exhibited in the wind tunnel and mean responses to different treatments in the field experiment were evaluated with a generalized linear model (GLM), modified for a Poisson distribution for the field data (Lo et al. 2013; Borrero-Echeverry et al. 2015; Reger et al. 2020), followed by Tukey's HSD test (SAS Institute, Cary, NC, USA). In all cases, $\alpha = 0.05$. Graphs were drawn using Graph Pad Prism 8 software (GraphPad Software, San Diego, CA, USA).

Results

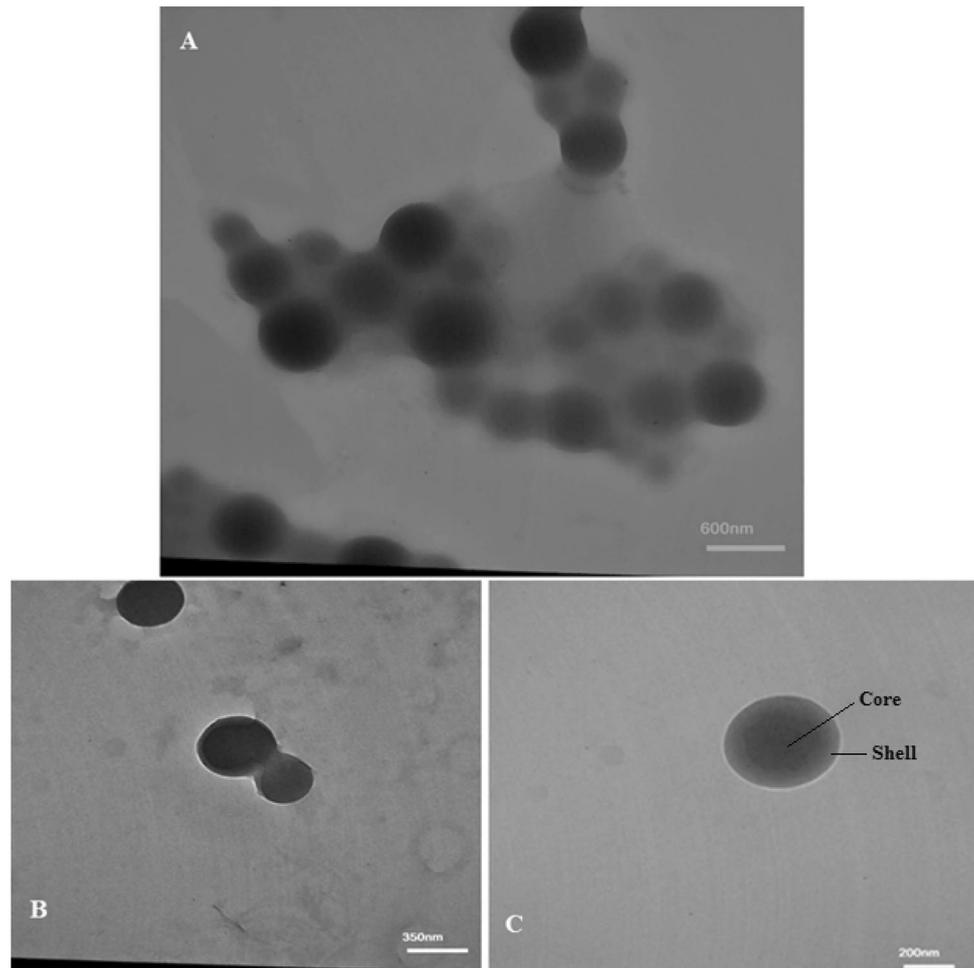
Nanocapsule characteristics

As shown in Fig. 2A–C, the nanocapsule trienal particles are relatively spherical and smooth, have a dense core and are partially in a gel. TEM images showed a size range of 300 to 350 nm, but the DLS spectra showed three distinct peaks in diameter (mean \pm SD), 184.2 ± 22.7 nm, 1109 ± 234.5 nm and 6293.4 ± 787.4 nm. Because of the agglomeration of nanocapsules, there are three peaks and a broader particle size distribution in DLS spectra (Fig. 3).

Wind tunnel tests

In the wind tunnel experiment, 80% of test male *E. ceratoniae* contacted the gel nanocapsule lure, compared to 70% for (*Z,E*)-7, 9, 11-dodecatrienyl format, and 60% for the unformulated trienal, and no males were attracted to the control. When exposed to the nanocapsule gel stimulus, males

Fig. 2 (A, B, C). Transmission electron microscopy images (TEM) of nanocapsules of carob moth sex pheromone (original) (Adapted from Danaye-Tous et al. 2020)



initiated flight in a mean time of 8.10 ± 0.57 s (mean \pm SE), slightly but significantly faster than 9.97 ± 0.49 s for (*Z,E*)-7,9,11-dodecatrienyl, while the take-off time for the unformulated trienal was not significantly different from either of the two ($F_{2,12} = 4.23$, $P = 0.04$, Fig. 4). There was no difference among treatments for the time spent in oriented flight ($F_{2,12} = 1.80$, $P = 0.20$), approaching ($F_{2,12} = 2.62$, $P = 0.11$) or sitting on the stimulus source ($F_{2,12} = 1.60$, $P = 0.24$) or in the total duration of the orientation sequence ($F_{2,12} = 0.98$, $P = 0.40$). However, males spent 29.47 ± 0.66 s attempting to copulate with the nanocapsule lure significantly more time than either of the other two stimuli ($F_{2,12} = 5.41$, $P = 0.02$).

Field tests

In the first week, the highest numbers of male *E. ceratoniae* (2.5 ± 0.29) were captured in traps baited with unformulated trienal; there was no significant difference between this catch

and that in traps baited with nanocapsules or (*Z,E*)-7,9,11-dodecatrienyl, which in turn attracted no more males than traps baited with the trienal formulated in glycerin or unbaited control traps ($F_{4,12} = 10.02$, $P < 0.001$, Fig. 5). After the first week, no males were captured in unbaited control traps. In weeks 2 and 3, the number of males captured by traps baited with trienal nanocapsules caught the highest numbers of males (2.75 ± 0.48 and 3.0 ± 0.41 , respectively), but in week 2 the catch in traps baited with nanocapsules was not significantly higher than in traps baited with unformulated trienal ($F_{4,12} = 12.57$, $P < 0.001$) and in week 3 it was no greater than in traps baited with (*Z,E*)-7,9,11-dodecatrienyl formate ($F_{4,12} = 13.15$, $P < 0.001$) (Fig. 5). In week 4, traps baited with trienal nanocapsules caught significantly more male *E. ceratoniae* (3.5 ± 0.29) than traps baited with (*Z,E*)-7,9,11-dodecatrienyl or the glycerin trienal formulation ($F_{4,12} = 50.71$, $P < 0.001$) (Fig. 5). In the fifth week, all traps caught very low numbers of males and there was no difference among treatments ($F_{4,12} = 1.31$, $P = 0.3219$) (Fig. 5).

Fig. 3 DLS spectra of nano-capsules of carob moth sex pheromone (original)

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13990707-Alizadeh-DLS-Size-Scan3.nsz
Measurement Results

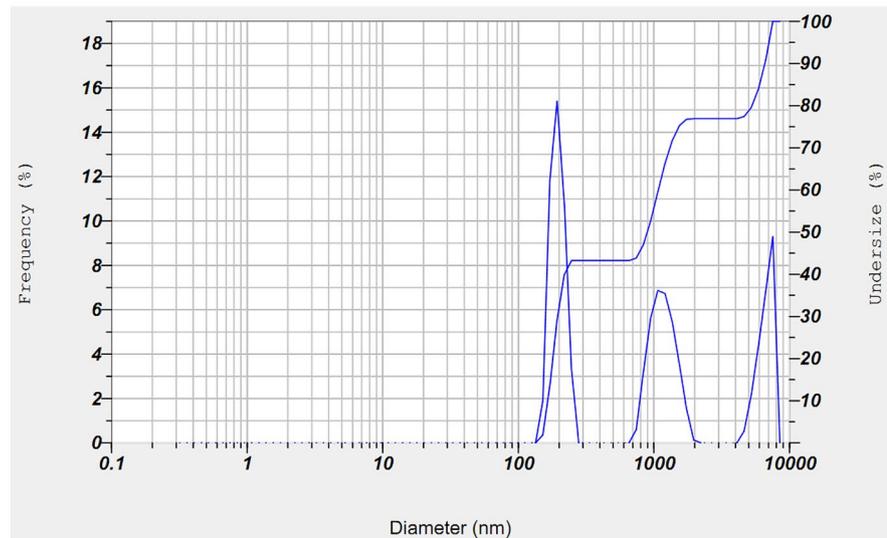
Date : Monday, September 28, 2020
 Measurement Type : Particle Size
 Sample Name : 13990707-Alizadeh-DLS-Size
 Scattering Angle : 90
 Temperature of the Holder : 25.1 °C
 Dispersion Medium Viscosity : 0.893 mPa·s
 Transmission Intensity before Meas. : 32208
 Distribution Form : Standard
 Distribution Form(Dispersity) : Polydisperse
 Representation of Result : Scattering Light Intensity
 Count Rate : 23 kCPS

Calculation Results

Peak No.	S.P.Area Ratio	Mean	S. D.	Mode
1	0.43	184.2 nm	22.7 nm	181.2 nm
2	0.34	1109.6 nm	234.5 nm	1015.2 nm
3	0.23	6293.4 nm	787.4 nm	6777.0 nm
Total	1.00	1908.5 nm	2471.6 nm	181.2 nm

Cumulant Operations

Z-Average : 2650.1 nm
 PI : 1.935



1/1

Fig. 4 Mean (\pm SE) times spent by male *E. ceratoniae* in a wind tunnel to take off, oriented flight, close approach, sitting, trying to mate and total time in different treatments. Bars with different letters for each behavior are significantly different; Tukey’s HSD test, $P < 0.05$

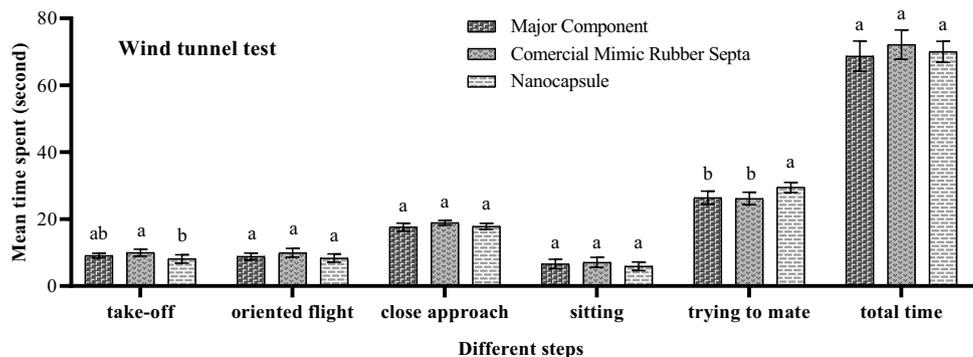
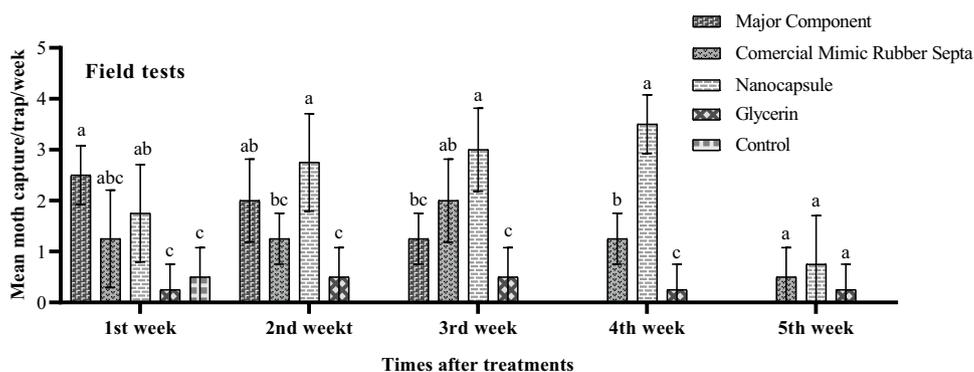


Fig. 5 Mean (\pm SE) captures of *E. ceratoniae* males per week for each treatment in a field trapping experiment from September 7 to October 11, 2019. Bars with different letters at each time point are significantly different, Tukey's HSD test, $P < 0.05$



Discussion

Our results showed that the nanocapsule gel is superior to other pheromone formulations in attracting male *E. ceratoniae* in a wind tunnel, that traps baited with trienal nanocapsules capture more males than traps baited with other pheromone formulations, and that the trienal nanocapsule gel formulation remains effective in attracting males for up to 4 weeks in the field. Therefore, we can assume that the amount of the remaining pheromone in the capsules in a longer time was still high enough and independently from the agglomeration that is a desired effect of the prepared formulation. The microencapsulated synthetic formulation in this study in addition to increasing the efficiency could be a suitable controlled release formulation, and nanocapsule gel formulation could have potential applications in mass trapping and mate disruption of carob moth in the field.

In this study, the nanocapsules were dispersed in three size ranges, with the largest mean diameter measuring 6,293.4 nm and accounting for 23% of the particle volume, while particles of mean diameters of 184.2 nm and 1,109.6 nm accounted for 43 and 34% of the volume, respectively (Fig. 3). The small particle size of the lower two categories, as well as the spherical shape of all particles, allows for optimal pheromone loading and controlled release (Perratti et al. 2013; Stadler et al. 2018; Heidary et al. 2022).

Our results support those of Baker et al. (1991), who also demonstrated initiation of flight by male *E. ceratoniae* in a wind tunnel in response to trienal alone. The high percentage (80%) of male *E. ceratoniae* exhibiting mate-seeking behavior in the wind tunnel to trienal nanocapsules compares very favorably with wind tunnel responses of 76.9–85.7% of *E. ceratoniae* males to used pheromone of three different populations (Ziaadini et al. 2011). It is superior to the 65% response of male *E. ceratoniae* flying toward an 8:1:1 blend of trienal, (*Z, E*)-9,11-tetradecadienal (dienal) and (*Z*)-9-tetradecenal (monoenal), a 63% response to the above blend with (*Z, E*)-7,9,11-dodecatrienyl formate replacing trienal and respective responses of 50 and 70% to trienal and (*Z, E*)-7,9,11-dodecatrienyl formate alone (Todd et al. 1992).

The pheromone component analogue (*Z, E*)-7, 9, 11-dodecatrienyl formate is reportedly not very effective as a field trap lure (Avand-Faghih et al. 2012; Dhoubi et al. 2016), but in our experiment it provided trap catches comparable to those in traps baited with trienal nanocapsules in weeks 1 and 3 (Fig. 5). Moreover, the catch rate of 0.17 males/trap/night is almost identical to the rate of 0.168 males/trap/night to traps baited with a commercial (*Z, E*)-7, 9, 11-dodecatrienyl formate lure (Naserian et al. 2016).

Our results showing superior performance of traps baited with unformulated trienal in week 1, and then a decline to zero catches in week 4 are consistent with initial high release of a highly volatile compound and its expected inactivation due to oxidation over time (El-Sayed et al. 2006, 2021). In contrast, traps baited with the trienal nanocapsule rose to a higher level than other treatments by week 2 and were at their highest level and significantly higher than to other traps in week 4 before declining to a very low level by week 5. The performance suggests a slow rise to an optimal controlled release rate and also that microencapsulation protected the trienal from oxidative degradation, as predicted by (Ziaee et al. 2014). The results suggest that further studies are warranted to develop a commercial microencapsulated pheromone lure for use in monitoring *E. ceratoniae* populations as a component of integrated pest management programs, and also that a microencapsulated formulation should be tested for effectiveness in mating disruption.

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